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TAT-02-F-05984

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# Sampling and Operations Plan for Asbestos Dump Keyersville, New Jersey As part of NPL Removal Assessment, 1990

Prepared for:
Nick Magriples
Removal Action Branch
U.S. EPA Region II
Edison, New Jersey 08837

Prepared by: Beverly Lawson

Region II Technical Assistance Team Roy F. Weston, Inc. Edison, New Jersey 08837 ABD 002 0084

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### Removal Action Branch U.S. EPA Region II Edison, NJ 08837

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Figure 1: Site Location Map

Figure 2: Sample Locations

- 1. PROJECT NAME: Asbestos Dump Meyersville Morris County, NJ
- 2. PROJECT REQUESTED BY: Nick Magriples
  Removal Action Branch
- 3. DATE REQUESTED: August 9, 1990
- 4. DATE OF PROJECT INITIATION: August 2, 1990
- 5. PROJECT ORGANIZATION AND RESPONSIBILITY:

The following is a list of key project personnel and their corresponding responsibilities:

Nick Magriples, USEPA Project Director

Beverly Lawson, TAT II Overall Project Coordination

Sampling Operations QC

Anibal Diaz, TAT II Laboratory Coordination & QC

6. PROJECT DESCRIPTION:

### A. Site Description

The Asbestos Dump site consists of four areas located in Morris County New Jersey which together comprise a National Priority List (NPL) site. Two of these areas; the New Vernon Road site and the White Bridge Road site are the focus of this sampling operation. A brief history of these areas follows.

The New Vernon Road site consists of approximately 30 acres of land off New Vernon Road, in Meyersville, New Jersey. The New Vernon Road site was operated as a corn and dairy cattle farm from 1945-1980. During the late 1960's, asbestos refuse consisting of loose asbestos fibers, broken asbestos tiles and sidings was landfilled in two areas of the site. A small depression in the westernmost section of the property was filled first. Then, a larger depression in the middle of the property was filled. Both areas were later graded and seeded. The New Vernon Road site contains asbestos wastes in a small landfill area in front of the private residence, in the main landfill area in the center of the property, along the dirt path that traverses north-south along the middle of the property, and in the area of the shed located next to the private residence. The thickness of the waste is not known.

The White Bridge Road site consists of about 12 acres also located in Meyersville, New Jersey. The site is bordered by the Great Swamp National Wildlife Refuge to the east and south and by private residences to the north and west. The site Was a farm until 1969, when the current owner conducted

### E. Parameter Table

#### \*. Aqueous Samples:

<u>Parameter</u>	Sample Watrix	Sample	Anely. Mth. Ref.	Sample	Holding <u>Time</u>	Limit of Detection	Volume	Preserv.
4spestos	Solia	•	TEM	4/A	N/A	4/4	50 g	N/A
Imass Cuttings	Solia	•	TEM	4/A	×/A	4/A	25 g	H/A
Dust Samples	Solid	•	TEN	N/A	N/A	N/A	25 g	N/A

<sup>\*</sup> The actual number will be based upon availability.

### 7. Sampling Procedures:

All soil samples will be collected between 0-6 inch depth using a trowel. The sample will then be transferred to a wide mouth 4 oz glass jar. The grass cuttings will be collected from the lawn mower using a trowel and transferred to a wide mouth 4 oz jar. The dust samples will be collected from the inside of a vacuum cleaner bag by pouring the samples directly into the sample jar. If this is not feasible, then the samples will be collected with a trowel and transferred to a wide mouth 4 oz jar.

### 8. <u>Sample Containers:</u>

All sample containers will be I-Chem laboratory precleaned glassware, as specified by the USEPA Sample Management Office Contract Lab Program.

### 9. <u>Sample Label:</u>

Each sample will be accurately and completely identified. All labels will be moisture resistant and able to withstand field conditions. Sample containers will be labeled prior to sample collection. The information on each label will include the following, but is not limited to:

- i. Date of collection
- ii. Site name
- iii. Sample identify/location
  - iv. Analysis requested

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### 14. Corrective Action:

All provisions will be taken in the field and laboratory to ensure that any problems that may develop will be dealt with as quickly as possible. This will be done to ensure the continuity of the sampling program. Any deviations from this sampling plan will be noted in the final report.

### 15. Reports:

Laboratory results and all requested QA/QC information will be submitted to EPA upon completion of sample analyses. Sampling reports will be issued after receipt of laboratory results.

### 16. Project Fiscal Information:

Sampling equipment and manpower shall be provided by the Technical Assistance Team (TAT) in coordination with the USEPA. All man-hours expended by TAT will be charged to TDD = 02-90-0113.

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Appendix A

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### Suite 201, 1090 King Georges Post Road. ■ Edison, NJ 08837, • (201) 225-6116 • FAX (201) 225-7037

#### TECHNICAL ASSISTANCE TEAM FOR EMERGENCY RESPONSE REMOVAL AND PREVENTION EPA CONTRACT 68-01-7367

TAT-02-F-05981

#### MEMORANDUM

Jean Williamson

International Asbestos Testing Laboratories

FRCM:

Andre' Bridgett, TAT II

CATE:

July 24, 1990

SUBJECT: Contractual Adreement for Analysis of Sambles from

Ascestos Dump for Aspestos

The purpose of this memo is to document our verbal agreement of July 14, 1990, concerning the analysis of twelve (12) soil samples for Aspestos. The samples will be delivered to you on or before August 3, 1990. The following commitment was made by IATL.

- The analysis of these samples will conform to TEM, which will include a breakdown by percentage of the different types of Aspestos.
- The total cost for the analysis will be \$3,600.00.
- The TA/QC will consist of performing and retaining a calibration and running as many suplicates as necessary to establish precision.
- Laboratory turnaround time for all samples submitted will be seven (7) days to written report. The written results will, therefore, be available at Weston no later than August 10, 1990. For delays, not due to acts of God, a reduction in the total charge will be 1 percent for the first day, an additional 2 percent for the second day and an additional 1 percent for each additional day.

If there are any questions concerning this memo, please contact me at 101) 115-6116.

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Rantan Camer 60 Fieldcrest Avenue Educa, NI 08837 2011 223-6040 Fax (201) 225-4577 July 25, 1990



Mr. Andre Bridgest ROY F. WESTON 1090 Kinz Georges Rosa Suite 20: Edison, rew Jersey 08837

Dear Mr. Bridgett:

Subject: Analytical Services for the Asbestos Dump Project

Confirming our relephone conversation of July 24, 1990. Clayton Environmental Consultants, inc. will provide analytical services to Roy F. Weston for the aspestos dump project as you requested. The following scope of work is based on the information you provided.

Roy F. Weston will submit 10 to 12 samples to Clayton on August 2, 1990. The samples will include soil surfaces, grass cumnes, and vacuumed dust. Clayton will perform transmission electron microscopy (TEM) analysis for the submitted samples. TEM is specific for aspestos and determines the percent and type aspestos fibers in a materiai sampie.

Clayton will provide these services using its commercially reasonable best efforts consistent with the level and skill ordinarily exercised by members of the profession currently practicing under similar conditions.

Clavion will provide the analytical services described herein at per sample. A 7-calendar-day numaround time will be provided for analytical results from the case samples are received.

Enclosed is a copy of our standard Terms and Conditions (Item A. Laboratory) [U.S.] - 11/88) which are incorporated in this confirmation letter as if set forth herein in full.

Thank you for the opportunity to provide our services to Roy F. Weston. Please call if you have any questions.

" ---- " nmarm . You, M . (duen, M . Wavne, M .

Sincereiv.

Odene B. Mina Technical Supervisor Laboratory Services

OBM/ee Encionine

MDE 0008481

July 24, 1990

Andre Bridgett
Roy F. Weston
1090 King George Post Road
Raritan Plaza Suite 210
Edison, NJ 08837

Dear Mr. Bridgett:

Thank you for choosing International Asbestos Testing Laboratories (IATL) for transmission electron microscopy analysis of your bulk samples.

TATL is a wholly owned subsidiary of Environmental Control Group (ECG), a public corporation with 1989 sales of \$77 million. Our full range of capabilities include phase contrast microscopy (PCM), polarized light microscopy (PLM), scanning electron microscopy (SEM), transmission electron microscopy (SEM), transmission electron microscopy (TEM) for the analysis of aspestos. Additionally, TATL now analyzes lead samples by atomic absorption spectrophotometry (AA).

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The professional staff at IATL maintains the highest standards in quality control and is fully accredited by the National Voluntary Laboratory Accreditation-Program (NVLAP). IATL also participates in the NIOSH Proficiency Analytical Testing Program (PAT) and the American Industrial Hygiene Association (AIHA) Asbestos Analyst Registry (AAR).

As we discussed on the telephone today, TATL will perform bulk sample analysis by transmission electron microscopy for your up coming project at [1] each. TATL vill provide one week turnaround for your twelve bulk samples with the certificate of analysis to be faxed and than hard copy to be mailed within three working days of completed analysis.

Should you have any questions regarding our laboratory, please do not hesitate to contact me. I look forward to working with you.

Sincerely,

Jean K. Williamson Laboratory Director D 002

July 24, 1990

Roy F. Weston, Inc. 1090 King Georges Post\_Road Suite 201 Edison, NJ 08837 (201) 225-6116

Attn: Andre Bridgett

Dear Mr. Bridgett:

Based on our conversation this afternoon, I have developed the following cost schedule:

)/sample

Bulk samples of soil, dust, and grass cuttings will be analyzed for asbestos content.

Samples will be pre-weighed, then desiccated overnight at approximately 400°F. The remaining sample will then be wasned with dilute HCl acid and placed in centrifuge. The final dry weight will be determined and TEM analysis will be performed for percentage and type of asbestos.

If you have questions, or require additional information, please contact me.

Very truly yours,

Tony Damato Program Manager Industrial Hygiene

<del>.</del>

TD/rk Proposal #0409

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MDE 0008483

# Precautionary Measures Against Hidden Hazards in Laboratory Samples

### Background

Under the authority of Section 104 of the Comprehensive Environmental Response. Commensation, and Liability Act (CERCLA or Supertuna) of 1980, Section 311 of the Clean Water Act, and Subtitle I of the Resource Conservation and Recovery Act (RCRA), EPA has been delegated the responsibility to undertake response actions with respect to the release or potential release of oil, petroleum, or harmfolds substances that pose a substance interact to human health or western, or the environment. In accition, EPA provides technical assistance to help mitigate encangerment of the public health, weifare or environment curing other emergencies and natural disasters.

EPA's successful implementation of these emergency response action responsibilities requires that technical support inpantities be provided in the form of contracted Technical Assistance Teams (TAT) for each EPA Region. The WESTON TAT Contract 68-01-7367 provides support to EPA Regions I. II. III. IV. V. ERT - Edison. and Hencounters - Washington.DC.

### Notice to Laboratory Management

The samples which will be shipped to your laboratory for analysis in accordance with applicable D.O.T. or IATA Requisitions have been collected by the WESTON TAT and have been tentatively designated by the field response than as either environmental or naturatous material samples.

in general. Environmental Samples are collected from streams, farm ponds, small lakes, wells, and off-site soils that are not reasonably expected to be contaminated with harmous materials. Samples of on-site soils of water, and materials collected from unims, bulk storage tanks, obviously contaminated ponds, impoundments, lagoons, pools, and leadnates from harmous waste sites are considered Harmous Samples. Samples which are obviously from a known radioactive material contamination site of which demonstrate beta of gamma activity greater than three times average background as scanned with a Geiger-Mueiler radiation survey meter are considered Radioactive Samples.

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Appendix B
TEM METHOD

2.2



### 2.2a TIM Water Sample Preparation Procedures

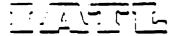
Background: Transfer of particulate from sampled water is essential. The object is to produce an intact carbon film containing the particles from the filter surface which is sufficiently clear for TEM analysis.

### Transfer of Suspended Particulate

- 1. A vacuum filtration system is utilized to collect the suspended particles in the sample.
- 2. Place a 0.5um filter on the holder to act a dispersion pad. On this place a 0.45um filter to catch the particles.
- 3. Remove a measured amount of the sample to be filtered. If the amount is less than 15ml. then add 20-30ml. of particle free water to the sample prior to filtering to ensure an even distribution of the specimen.
- 4. Slowly introduce the measured sample to the filter funnel as to not damage the filter. Swirl the funnel to ensure that the specimen is homogenized.
- 5. Allow to settle for a moment before introducing vacuum. Apply suction until the sample has been filtered completely. (DO NOT RINSE THE FUNNEL WALLS WILE THE FILTER IS IN PLACE THIS WILL DISTURB THE SAMPLE DISTRIBUTION)
- Do a series of dilutions. ie: 1mm, 5mm, 10mm etc.
   Allow the filter minus the diffusion pad to dry thoroughly under NEGATIVE air.

SEE MCE FILTER PREP. TO COMPLETE PROCEDURES

ABD 002 010;



### 2.2c TEM Solid Sample Preparation Procedures

Background: Homogenized breakdown of solid specimen to dissociate asbestos particulate. The object is to produce an intact carbon film containing the particles from the specimen which is sufficiently clear for TEM analysis.

### Acquisition of Fibrous Particulate

### Treatment il: (Allows for Percent Estimation)

- 1. Bulk-solid samples are gently and slowly ground to a powder.
- 2. A weighed portion is then suspended in a known volume of particle free water and briefly ultrasonicated to disperse the particulate.
- 3. The solution is then placed into a volumetric flask and brought to volume with particle free water.

Treated as a water sample.
(See Section 2.2b Water Sample Preparation)

# Treatment #2: (Does Not Allow for Percent Estimation)

- 1. Weighed shaved cross sections of the sample are placed onto a clean watch glass.
- 2. Tetrahydrofuran (THF) is then slowly piperted onto the sample to carefully dissolve the surrounding matrix material.
- 3. After the THF has sufficiently dried the sample is then oxygen plasma ashed for 15 minutes.
- 4. The sample is then placed into 100ml. of particle free water.

Treat as a water sample. (See Section 2.2b Water Sample Preparation)

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Appendix C
Soil Sampling SOPs

### SOIL SAMPLING SOP

This recommended protocol outlines procedures and equipment for the collection of representative samples from surface and subsurface locations.

Surface sampling commonly refers to the collection of samples at a C-6 inch depth. This is most efficiently accomplished with the use of a trowel or scoop. For samples at lower depths, a decontaminated bucket auger or power auger may be needed to advance the hole to the point of collection. Another clean bucket auger can then be used to collect the sample. For samples at depths greater than three feet, the use of a drill rig and split spoon sampler will be necessary. In some situations, sample locations can be accessed with the use of a backhoe.

Whether surface or subsurface, and whether a bucket auger or drill rig is used to access the sample, several considerations are important during soil sample collection. An attempt must be made to maintain sample integrity by preserving its physical form and chemical composition to as great an extent as possible. First, the mechanism used to advance the hole must be properly decontaminated. The device then used for actual sample collection should not be same as that used to advance the hole. This instrument should be appropriately decontaminated, as should any instrument utilized to transfer the sample into the sample bottle.

Secondly, care must be taken in handling the sample. The sample should be transferred into the sample bottle as quickly as possible, with no mixing, to assure that the volatile fraction is not lost. It is also recommended that for volatile organics analysis of soils, the laboratory performing the analysis should provide wide mouth bottles (4 ounce) for sample collection. This will reduce disturbance of the sample and may help prevent the loss of volatiles.

Soil sampling is generally accomplished through the use of one of the following samplers:

- scoop or trowel
- tulip bulb-planter
- bucket auger
- soil coring device/silver bullet sampler
- waste pile sampler
- power auger (in conjunction with another device)
- split spoon sampler
- Shelby tube sampler